universitat wien Variable enzymatic activity of prokaryotes under different hydrological settings in a river floodplain system of the Danube

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Introduction

River-floodplain systems play a significant role in organic matter cycling and carbon mineralization, sequestration and transport (Battin et al. 2008). In floodplains, the driving force of lateral exchange processes is hydrological connectivity, both via surface flow and via groundwater inflow (Tockner et al. 1999). A previous study revealed that hydrological connectivity between the main channel and a floodplain segment influenced productivity of both free-living (FL) and particle-associated bacteria (PA) (Luef et al. 2007). There is also evidence that hydrological retention can influence hydrolytic extracellular enzymatic activity (EEA) in floodplain sediments (Burns and Ryder, 2001), however the activity of oxidative enzymes may show no relation to hydrology (Williams, 2000). The extent to which FL and PA enzymes contribute to overall activity may be highly variable as a result of biological and physical processes, such as variable hydrological connectivity. However the question if variable hydrology, especially frequency of connection is mirrored in enzymatic activity in river floodplain systems remains largerly unknown.

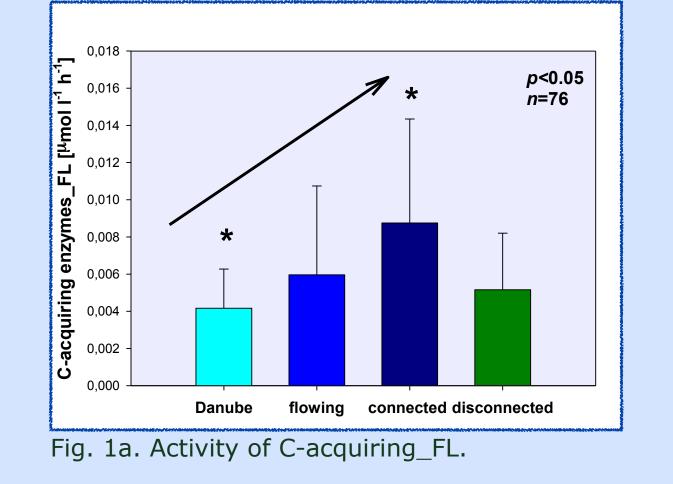
Hyphothesis

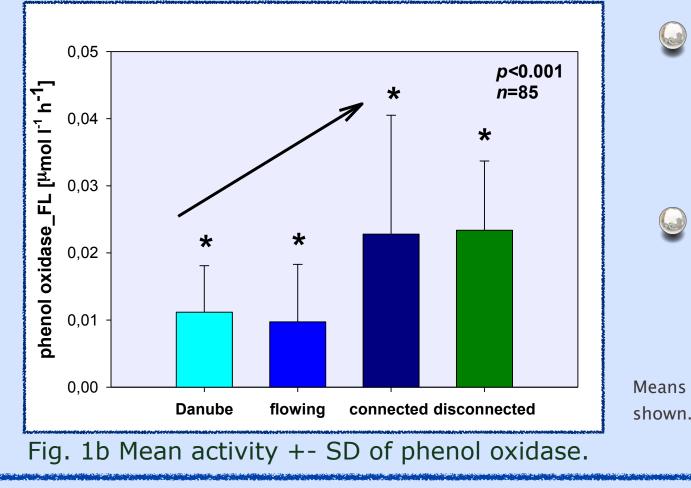
Degree of connectivity and water exchange in floodplain lakes influences prokaryotic enzymatic activity both free-living (FL) and particle-associated (PA).

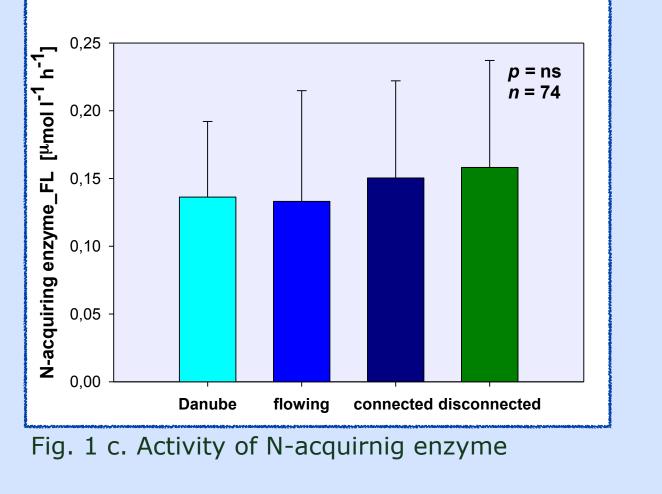
Results and discussion

Enzymatic acctivity in ambient water (FL) under different hydrological connectivity

Enzymatic acctivity associated with particles (PA) under different hydrological connectivity

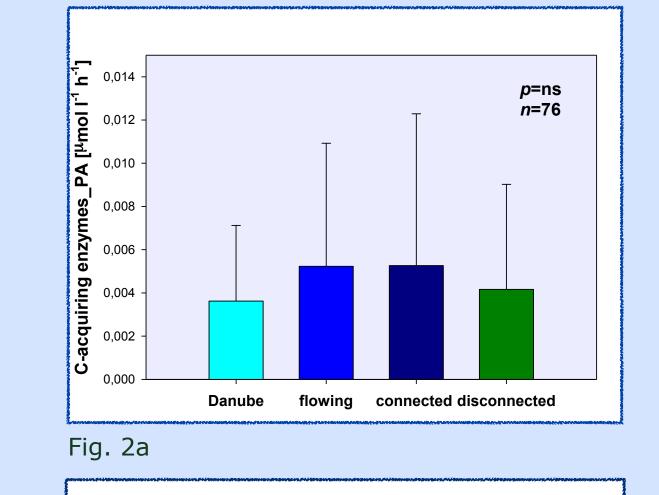


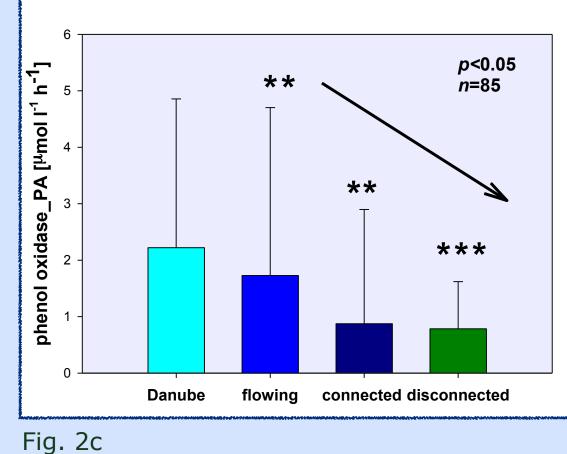


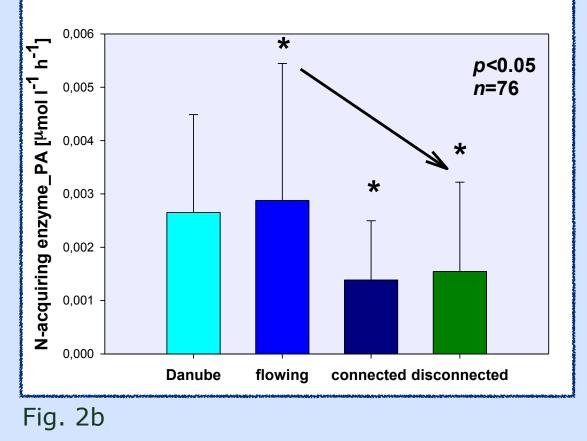


- Activity of C-acquiring enzymes and phOx in ambient water (FL) increased together with decreasing connectivity (Fig.1a,b).
- N-acquiring enzymes did not show clear dependencies on the degree of connectivity (Fig. 1c).

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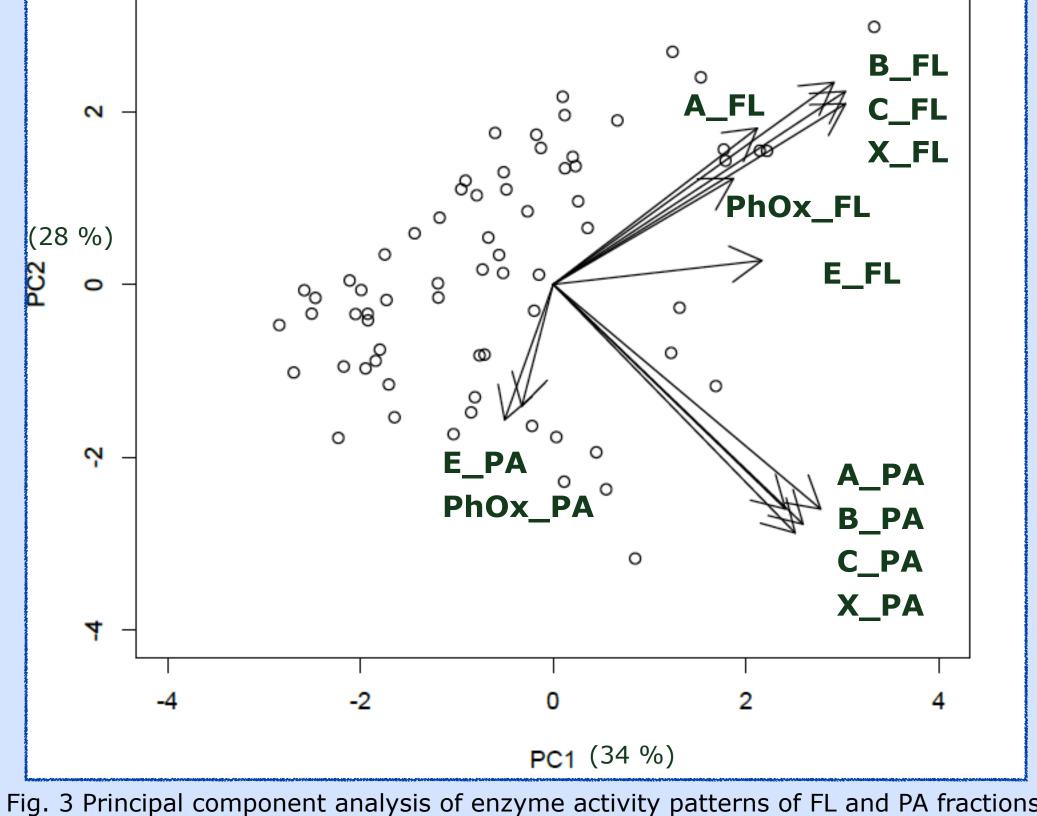






Solution Along with a gradient of connectivity, no significant changes in activity of C-acquiring_PA enzymes was observed (Fig.2a). Sectivity of N-acquiring enzyme and phenol oxidase associated with particles (PA) decreased with the degree of connectivity (Fig.2b,c).

Conclusions Particle-associated enzymes and free-living enzymes are strongly **B_FL** interlinked what Hydrological connectivity of • Of all the measured enzymes, C FL suggests a tight coupling 8 X_FL floodplain lakes with the main PhOx_FL showed the most clear 00 within each fraction. 80 channel is important factor which dependency on a gradient of PhOx_FL drives microbial enzymatic activity. connectivity (r=-0.43 p<0.0001), E_FL 0 000 Decreasing connectivity promotes (Fig.1b). This implies that activity of FL enzymes, whereas prolonged residence time in Solution The strong correlation activity of particle-associated between PhOx_FL and E_FL floodplain lakes allows more during flowing conditions enzymes declines with decreasing refractory material to be utilized. A_PA (r=0.77,p<0.01) and connectivity. PhOx PA



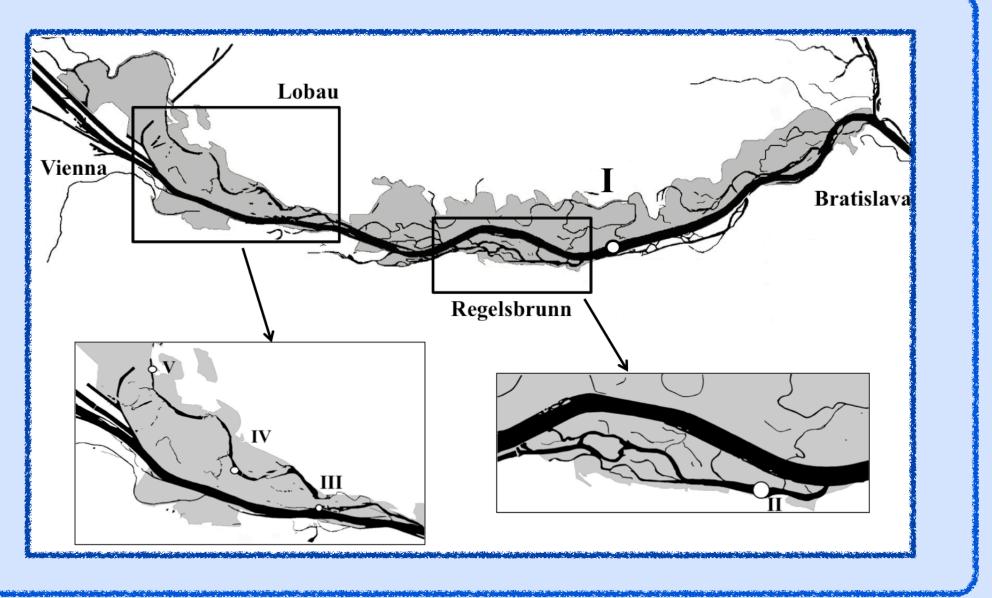
FL: free-living fraction PA:particle-associated fraction

E_PA and PhOx_PA during disconnected conditions (r=0.67,p<0.0001) suggests a tight coupling of enzymes responsible for utilization of labile and refractory material.

In dynamic river-floodplain systems, under variable hydrological settings, there is opportunity for enzymatic degradation of refractory material in the additional presence of enzymes responsible for utilization of semi-labile organic matter.

Study site

Sampling locations in the Danube riverfloodplain near Vienna (Austria) which represent a gradient of connectivity to the main river.



Methods Bacterial parameters Section [m] (Peduzzi Sector Extracellular enzymatic activity (EEA) DOM quantity and quality • C-acquiring enzymes:-β-d-glucosidase, • DOC [mg l⁻¹] (Benner and Strom 1993) a-d-glucosidase, β -d-xylosidase, • DOM origin: FI ratio (McKnight et al. cellobiohydrolase • Molecular size indicator: •N-acquiring enzyme:endopeptidase Slope ratio Helms 2008) (Hoppe, 1993) phenol oxidase (phOx) (Pind et al. 1994) •Aromaticity: SUVA₂₅₄ (Weishaar et al. 2003) • Abundance (Porter and Feig 1980) Chlorophyl *a* (Lorenzen, 1967) • Bacterial secondary production (Fuhrman and Azam, 1982) Selected references

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